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Preparation of 2- and 4-(2-alkylcarbamoyl-1-methylvinyl)-7-alkyloxybenzo[*b*]furans having potent antagonistic activity against human leukotriene B₄ BLT₁ and/or BLT₂ receptors[†]

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(*E*)-2-Acetyl-4-(2-diethylcarbamoyl-1-methylvinyl)-7-(1phenylethoxy)benzo[*b*]furan (4b) with a characteristic conformation and (*E*)-2-(2-morpholinocarbo-1-methyl-vinyl)-7-ethoxycarbopropoxybenzo[*b*]furan ((*E*)-3b) were prepared and evaluated for their leukotriene B₄ (LTB₄) antagonistic activity. Compound 4b showed potent antagonistic activity against human BLT₁ and BLT₂ receptors. Compound (*E*)-3b displayed selective BLT₂ receptor antagonistic activity. Both compounds were inactive to cysteinyl LT receptors.

Leukotriene B_4 (LTB₄) plays important roles in the host defence system against infection and invasion of foreign bodies, however overproduction of LTB₄ is involved in various inflammatory diseases.¹ Thus, many attempts have been made to develop antagonists of LTB₄ as clinical drugs. Of particular interest are ONO-4057,^{2a} ZK-158252,^{2b} BIIL315^{2c} and LY 293111^{2d} (Fig. 1).

These antagonists may be grouped into two classes, aliphatic carbon chain types (ONO-4057 and ZK-158252) and ether types (BIIL315 and LY293111). No LTB₄ antagonist has yet been marketed in spite of the clinical application of an LTD₄ receptor antagonist (cysteinyl LT₁ receptor antagonist). Recently, a second LTB₄ receptor (BLT₂) was discovered and its molecular cloning was reported.³ Current work on LTB₄ and its receptors

†Electronic supplementary information (ESI) available: Experimental details for compounds **3a**, **3b**, **3f**, **4b** and **4c**. See http://www.rsc.org/suppdata/ob/b4/b411286e/

suggest that LTB₄ selective antagonists may be applicable for treatment of arteriosclerosis,⁴ rheumatoid arthritis⁵ and pancreatic cancer,⁶ as well as for immunosuppression^{2c,7} of allograft rejection in organ transplantation. This encouraged us to find novel BLT₂ selective and dual BLT₁/BLT₂ selective receptor antagonists. BLT₂ selective antagonists may be also useful for clarifying the bioactive roles of the BLT₂ receptor in the body.

Simple heteroaromatic skeleton compounds of stable conformation may well be favored over the aliphatic carbon chain type and the ether type compounds.8 Benzopyran8b,8c and dibenzofuran^{8d} derivatives have been reported previously as LTB_4 antagonists. Here we used the benzo[b]furan derivative to search for novel LTB4 antagonists. The combination of conjugated triene and a single C=C bond near one OH group of possible LTB4 conformers (A, B) was based on consideration of the benzo[b]furan ring having an O(CH₂)₃COOR group originating from the partial structure of LTB₄. The α , β unsaturated carbamoyl group9 modified from cinnamic acid proved to be an interesting functional group showing cysteinyl LTs antagonistic activity in our recent study.¹⁰ Several 2alkylcarbamoyl-1-methylvinyl groups¹¹ devised from the α,β unsaturated carbamoyl group were introduced at C-2 or C-4 of the benzo[b]furan ring. Introduction of functional groups at C-2 or C-4 is preferable to other positions to examine the relationship between the substituent position and bioactivity because the C-2 and C-4 positions have significantly different stereochemical and electronic environments. We report here the



structural characteristics of I and II and their LTB_4 antagonistic activity characterized by the potency and selectivity for human BLT_1 and/or BLT_2 receptors (designed compounds I and II, Fig. 2).



Fig. 2 Possible conformers (A, B) of LTB₄ and designed benzo[*b*]furan derivatives (I, II).

Alkylation of 2-acetyl-7-hydroxybenzo[b]furans (1a, 1b) with alkyl halides gave the 7-alkyloxybenzo[b]furans (2a, 2c-2e). Treatment of 2a with phosphonoamides (5)¹² under Horner-Wadsworth-Emmons (HWE) reaction13 conditions afforded (E/Z)-2-(2-alkylcarbamoyl-1-methylvinyl)benzo[b]furans ((E/ Z)-3a, -3b, -3c). Compounds 2b, 2c and $2f^{14}$ were also subjected to HWE reaction with the phosphonoamide (5) to give (E/Z)-2-(2-alkylcarbamoyl-1-methylvinyl)benzo[b]furans ((E/Z)-3d, -3e and -3f), respectively. Yields and ratios of E/Z of 3 are shown in Scheme 1. (E)-Isomers ((E)-3a, -3b, -3c, -3e, -3f) were isolated from the corresponding E/Z mixtures ((E/Z)-3a, -3b, -3c, -3e, -3f). The (E)-isomers (E)-3 showed only a nuclear Overhauser enhancement (NOE) correlation between the olefinic CH₃ and 3-H. This could be explained reasonably, since the double bond of the (E)-2-(2-alkylcarbamoyl-1-methylvinyl) group had an strans configuration with a double bond between C-2 and C-3, and also this group lay on approximately the same plane as the benzo[b]furan ring.¹⁵ The stereostructure of (E)-3f, as a representative compound of (E)-**3**, was determined by X-ray analysis as shown in Fig. 3,¹⁶ with the torsion angle being 6.9°.¹⁵ Conformation and NOE correlation of (E)-**3b** are shown in Fig. 4. The main isomer of (E/Z)-**3d** was isolated and identified as s-*trans*-(Z)-isomer, (Z)-**3d**, on the basis of its ¹H-NMR data.

Compounds **2c**, **2d** and **2e** were treated with 2-butenamides (6) in the presence of $(CH_3COO)_2Pd$, $(2-CH_3C_6H_4)_3P$ and $(C_2H_5)_3N$ under the Heck coupling conditions^{11b,17} to afford (*E*)-2acetyl-4-(2-alkylcarbamoyl-1-methylvinyl)-7-alkyloxybenzo[*b*] furans (**4a–4d**) on the basis of their NOE analyses (Scheme 1).

In the NOE of compounds 4, it was very interesting that both the olefinic CH₃ and olefinic H had NOE correlations with 3-H and 5-H, respectively. Consideration of the NOE correlations led to speculation concerning the molecular dissymmetry of 4 arising from restriction of the rotation about the single bond at the C-4.18 However, this speculation was rejected by the absence of the diastereoisomer of 4b which contained one asymmetric carbon atom in the substituent group at C-7 on the basis of ¹H-NMR studies. The torsion angle of representative compound 4c between the benzo[b]furan plane and the 2-alkylcarbamoyl-1methylvinyl group at C-4 estimated using MM215 was 44.7°. The X-ray structure of 4c is shown in Fig. 3,¹⁶ with the torsion angle being 45.7°. Conformation and NOE correlation of 4c are shown in Fig. 4. The 2-alkylcarbamoyl-1-methylvinyl groups of compounds (E)-3 and 4 showed distinctly different conformations from each other, because the 2-alkylcarbamoyl-1-methylvinyl group of compound 4 was subjected to significant steric hindrance from 3-H and 5-H, while there was little hindrance from 3-H on compound (E)-3 (Fig. 4).

Compounds ((*E*)-**3**, **4**) were evaluated for their LTB₄ antagonistic activity by two *in vitro* methods: Method A,¹⁹ inhibition of LTB₄-induced TXB₂ release from bronchoalveolar eosinophils of guinea pigs and Method B, inhibition of calcium mobilization in CHO-humanBLT₁ (CHO-hBLT₁) and CHO-humanBLT₂ (CHO-hBLT₂) cells by LTB₄.

Method A. Compounds (*E*)-3a, -3b, -3c, -3d, -3e, 4a, 4b, 4c and 4d at 100 μ M completely inhibited LTB₄ (100 nM)-induced TXB₂ release from the bronchoalveolar eosinophils. Compounds (*E*)-3a, -3b and 4b even at 1 μ M produced complete inhibition, but 4c showed 84% inhibition at the same concentration. Furthermore, the concentration-dependent antagonistic activity for (*E*)-3a, -3b and 4b was evaluated at the concentrations of 0.1 nM, 1 nM, 10 nM, 100 nM and 1 μ M. The most potent compound (*E*)-3a





Fig. 3 X-Ray structures of (E)-3f and 4c.



Fig. 4 Conformation and NOE correlations of (*E*)-3b and 4c.

showed almost complete inhibition at 100 nM and 1 μ M, and 26.8% inhibition at 10 nM. Compounds (*E*)-**3b** and **4b** were less active than (*E*)-**3a**: 18.7 and 17.1% inhibition at 10 nM, and 37.7 and 31.7% inhibition at 100 nM, respectively (Fig. 5).²⁰



Fig. 5 Effect of (*E*)-**3a**, (*E*)-**3b**, **4b** on LTB₄-induced TXB₂ release from bronchoalveolar eosinophils harvested from Sephadex G-200-treated with guinea pigs (mean \pm S.E., n = 3). (*E*)-**3a**, (*E*)-**3b** and **4b** was added 5 min before eosinophil stimulation by LTB₄ (100 nM). Statistically significant differences from the control are indicated (**P* < 0.05, ****P* < 0.001, Bonferroni's multiple test). Spon: Spontaneous, Cont: Control, a) Inhibition (%).

Method B. Three compounds ((*E*)-3a, -3b, 4b) selected from the findings with Method A were evaluated by Method B at concentrations of 10 nM, 100 nM, 1 μ M and 10 μ M,

in comparison with standard LTB₄ antagonists (ZK-158252 and ONO-4057). Compound **4b** showed the most potent and concentration-dependent inhibition of calcium mobilization in both CHO-hBLT₁ and CHO-hBLT2 cells and its potency lay between ZK-158252 and ONO-4057. Characteristically, (*E*)-**3b** showed selective and concentration-dependent inhibition in CHO-hBLT₂ cells (Fig. 6).²¹ In contrast, (*E*)-**3a** was found to be inactive by Method B.





Fig. 6 Effect of (*E*)-**3a**, (*E*)-**3b**, **4b** on calcium mobilization by LTB₄ (100 nM) in CHO-hBLT1 and CHO-hBLT2 cells (mean \pm S.D., n = 3). Statistically significant differences from the control are indicated (**P* < 0.05, ***P* < 0.01, unpaired *t*-test).

To prove that inhibition of calcium mobilization was not due to a simple cytotoxic effect, cytotoxicity studies using ATP were performed.²² Cysteinyl LT antagonistic activity assay²³ for the active compounds ((E)-3a, -3b, 4b) was also carried out to examine their selectivity for LTB4 antagonistic activity, and they were found to be inactive. Compound 4b, shown to be moderately active by Method A was revealed to be the most potent compound with greater potency than the standard compound ONO-4057 using both human BLT_1 and BLT_2 in Method B. Unfortunately, the most active compound (E)-3a according to Method A was completely inactive when examined by Method B. Thus, the substituent position (at C-4) and the conformation of the 2-alkylcarbamoyl-1-methylvinyl group described above for the structure of 4 may contribute to its antagonistic potency against human BLT receptors. On the other hand, (E)-3b with the 2-morpholinocarbo-1-methylvinyl group at the C-2, which lies on nearly the same plane as the benzo[b]furan ring showed selective activity for human BLT₂. Compound (E)-3b is, to the best of our knowledge, the first antagonist showing selective BLT₂ antagonistic activity.

In this study, we found a potent human BLT_1 and BLT_2 receptor antagonist, (*E*)-2-acetyl-4-(2-diethylcarbamoyl-1-methylvinyl)-7-(1-phenylethoxy)benzo[*b*]furan (**4b**), and a BLT_2 selective antagonist, (*E*)-2-(2-morpholinocarbo-1-methylvinyl)-7-ethoxycarbopropoxybenzo[*b*]furan ((*E*)-**3b**). The next step would be to synthesize new derivatives and evaluate them to find more potent and selective (2-alkylcarbamoyl-1-methylvinyl)benzo[*b*]furan derivatives. Such work should clarify the relationships between the selective antagonist activities and the stereochemistry of the functional groups in a series of these derivatives.

Experimental

(*E*)-2-(2-Morpholinocarbo-1-methylvinyl)-7-ethoxycarbopropoxybenzo[*b*]furan ((*E*)-3b)

To a suspension of NaH (60% in oil, 0.24 g, 6.1 mmol) in anhydrous THF (10 ml) was added dropwise a solution of [2-(4-morpholinyl)-2-oxoethyl]phosphonic acid diethyl ester (1.6 g, 6.2 mmol) in anhydrous THF (10 ml) under N₂ atmosphere at -5 °C with stirring. The solution was then stirred at 25 °C until it became clear. A solution of **2a** (1.0 g, 3.4 mmol) in anhydrous THF (15 ml) was added dropwise at 25 °C, and the mixture was stirred at 25 °C for 3 h. The reaction mixture was worked up by an ordinary procedure to obtain a residue which was purified by silica gel column chromatography [CHCl₃–ethyl acetate (10 : 1)] to give (*E*)-**3b** (0.34 g, 24.6%) as colorless needles. Mp 90.9–94.8 °C.

(*E*)-2-Acetyl-4-(2-diethylcarbamoyl-1-methylvinyl)-7-(1-phenyl-ethoxy)benzo[*b*]furan (4b)

A mixture of **2d** (1.0 g, 2.8 mmol), (*E*)-*N*,*N*-diethyl-2butenamide (0.47 g, 3.3 mmol), palladium acetate (0.031 g, 0.14 mmol), tri-*o*-tolylphosphine (0.085 g, 0.28 mmol) and Et₃N (10.0 ml, 0.072 mol) was heated at 90–100 °C for 16 h. The precipitate was dissolved with ethyl acetate, and the insoluble portion was filtrated off. The filtrate was evaporated to dryness. The residue was poured into ice water, then made acid with 5% HCl solution and extracted with ethyl acetate. The organic layer was washed with brine and dried. The solvent was evaporated off, and the resulting residue was purified by silica gel column chromatography [hexane–ethyl acetate (5 : 1)] to give a yellow solid. The solid was recrystallized from hexane–ethyl acetate to give **4b** (0.57 g, 48.7%) as yellow prisms. Mp 118.4-121.5 °C.

Measurement of calcium mobilization in CHO cells

CHO cells stably expressing human BLT_1^{24} and BLT_2^{3e} seeded on 96-well glass-bottom plate (Coster 3603) at 4×10^4 cells per well, were loaded with 4 μ M Fluo-3 (Dojin, Kumamoto, Japan) in 1 × HBSS (Hanks balanced salt solution, Sigma) containing 0.04% pluoronic acid and 1% FCS at 37 °C for 1 h. The cells were washed twice with 1 × HBSS and pretreated with various concentrations of antagonists diluted in 100 μ l of 1 × HBSS, 1% FCS for 30 min. A stock of BLT antagonists was prepared as a DMSO solution, and the final concentration of DMSO in the assay was adjusted to 0.1% in all wells. To each well was added 50 μ l of 300 nM LTB₄ (Cayman Chemicals) to give a final concentration of 100 nM, and the LTB₄-dependent increase in the fluorescent intensity was measured using FlexStation (Molecular Devices). CHO cells transfected with empty vector did not respond to 100 nM LTB₄ (data not shown).

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References

- (a) C. R. Turner, R. Breslow, M. J. Conklyn, C. J. Andresen, D. K. Patterson and A. Lopez-Anaya, J. Clin. Invest., 1996, 97, 381–387;
 (b) P. Sharon and W. F. Stenson, Gastroenterology, 1984, 86, 453– 460; (c) A. Nakao, K. Nasaka, N. Ohishi, E. Noiri, T. Suzuki and S. Taniguchi, Kidney Int., 1997, 63, 236–238; (d) R. J. Griffiths, E. R. Pettipher, K. Kock, C. A. Farrell, R. Breslow and M. Conklyn, J. Proc. Natl. Acad. Sci. USA, 1995, 92, 517–521; (e) R. P. Gladue, L. A. Carroll, A. J. Milici, D. N. Scampoli, H. A. Stukenbrok and E. R. Pettipher, J. Exp. Med., 1996, 183, 1893–1898.
- 2 (a) K. Kishikawa, S. Nakao, S. Matsumoto, K. Kondo and N. Hamanaka, Adv. Prostaglandin Thromboxane Leukotriene Res., 1995, 23, 279-281; (b) K. Kishikawa, N. Tateishi, T. Maruyama, R. Seo, M. Toda and T. Miyamoto, Prostaglandins, 1992, 44, 261-275; (c) H. Morita, K. Takeda, H. Yagita and K. Okumura, Biochem. Biophys. Res. Commun., 1999, 264, 321-326; (d) F. W. Birke, C. J. Meade, R. Anderskewitz, G. A. Speck and H.-M. Jennewein, J. Pharmacol. Exp. Ther., 2002, 297, 458-466; (e) R. D. Pallavi, K. H. Abdelmadjid, P. Mai, D. S. Wolf-Dieter, M. S. Bruce and W. Walter, J. Biol. Chem., 1999, 274, 23341-23348; (f) M. Matousek, K. Mitsube, M. Mikuni and M. Brannstrom, Mol. Hum. Reprod., 2001, 7, 35-42; (g) W. T. Jackson, L. L. Froelich, R. J. Boyd, J. P. Schrementi, D. L. Saussy, Jr., R. M. Schultz, J. S. Sawyer, M. J. Sofia, D. K. Herron, T. Goodson, Jr., D. W. Snyder, P. A. Pechous, S. M. Spaethe, C. R. Roman and J. H. Fleisch, J. Pharmacol. Exp. Ther., 1999, 288, 286-294; (h) K. Kuwabara, K. Yasui, H. Jyoyama, T. Maruyama, J. H. Fleisch and Y. Hori, Eur. J. Pharmacol., 2000, 402, 275-285
- 3 (a) C. Brink, S.-E. Dahlen, J. F. Evans, D. W. P. Hay, S. Nicosia, C. N. Serhan, T. Shimizu and T. Yokomizo, *Pharmacol. Rev.*, 2003, **55**, 195–227; (b) A. Toda, T. Yokomizo and T. Shimizu, *Prostaglandins Other Lipid Mediators*, 2002, **68-69**, 575–585; (c) T. Yokomizo, K. Masuda, K. Toda, T. Izumi and T. Shimizu, *Am. J. Respir. Crit. Care. Med.*, 2000, **161**, S51–55; (d) T. Yokomizo, T. Izumi and T. Shimizu, *Arch. Biochem. Biophys.*, 2001, **385**, 231–241; (e) T. Yokomizo, K. Kato, K. Terawaki, T. Izumi and T. Shimizu, *J. Exp. Med.*, 2000, **192**, 421–431.
- 4 (a) R. J. Aiello, P. A. Bourassa, S. Lindsey, W. Weng, A. Freeman and H. J. Showell, *Arterioscler. Thromb. Vasc. Biol.*, 2002, 22, 443–449; (b) A. Mennander, S. Tiisala, J. Ustinov, A. Räisänen, T. Paavonen and P. Häyry, *Arteriosclerosis and Thrombosis*, 1992, 12, 1380–1386.
- 5 (a) R. Alten, E. Gromnica-Ihle, C. Pohl, J. Emmerich, J. Steffgen, R. Roscher, R. Sigmund, B. Schmolke and G. Steinmann, *Ann. Rheum. Dis*, 2004, **63**, 170–176; (b) A. Hashimoto, H. Endo, I. Hayashi, Y. Murakami, H. Kitasato, S. Kono, T. Matsui, S. Tanaka, A. Nishimura, K. Urabe, M. Itoman and H. Kondo, *J. Rheumatol.*, 2003, **30**, 1712–1718.
- 6 W.-G. Tong, X.-Z. Ding, R. Hennig, R. C. Witt, J. Standop, P. M. Pour and T. A. Adrian, *Clin. Cancer Res.*, 2002, **8**, 3232–3242.
- 7 (a) H. Takatsuka, Y. Takemoto, S. Yamada, T. Wakae, A. Mori, M. Okada, N. Iwata and T. Okamoto, *Drugs Exp. Clin. Res.*, 2002, 28, 121–125; (b) M. Tanaka, T. Tamaki, Y. Konoeda, Y. Uchida, T. Kaizu and A. Kawamura, *Transplant Proc.*, 2000, 32, 2340.
- 8 Several heteroaromatic compounds as LTB₄ antagonist were reported previously as follows. (a) D. Delorme, Y. Ducharme, C. Brideau, C.-C. Chan, N. Chauret, S. Desmarais, D. Dube, J.-P. Falgueyret and R. Relean, J. Med. Chem., 1996, **39**, 3951–3970 (L 708780, naphthalene derivative with furan ring);; (b) S. W. Djuric, S. H. Docter, S. S. Yu, D. Spangler, B. S. Tsai, C. P. Anglin, T. S. Gaginella,

J. F. Kachur and R. H. Keith, Bioorg. Med. Chem. Lett., 1994,
4, 811–816 (SC-53228, benzopyran derivative);; (c) S. W. Djuric,
P. W. Collins, P. T. Jones, R. L. Shone, B. S. Tsai, D. J. Fretland,
G. M. Butchko, D. Villani-Prince and R. H. Keith, J. Med. Chem,
1989, 32, 1145–1147 (SC 41939, benzopyran derivative);; (d) T. S.
Shoupe, S. M. Coutts, D. C. Baker, E. S. Hand, Can. Pat. Appl., 1991
63pp. CPXXEB CA 2013960, AA 19910319, CAN 115:158949, AN
1991: 558949 (PF 10042, dibenzofuran derivative); (e) C. D. Wright,
P. J. Kuipers, M. D. Hoffman, D. O. Thueson and M. C. Conroy,
Biochem. Biophys. Res. Commun., 1990, 167, 828–834 (CI 949, indole
derivative); (f) J. M. Musser, U. R. Chakraborty, S. Sciortino, R. J.
Gordon, A. Khandwala, E. S. Neiss, T. P. Pruss, R. V. Inwegen, I.
Weinryb and S. M. Coutts, J. Med. Chem, 1997, 30, 96–104 (PF 5901,
quinoline derivative); (g) C. Can, M. G. Cinar, S. Ulker, A. Evinc and
S. Kosay, Eur. J. Pharmacol., 1998, 350, 223–228.

- 9 H. Nakai, M. Konno, S. Kosuge, S. Sakuyama, M. Toda, Y. Arai, T. Obata, N. Katsube, T. Miyamoto, T. Okegawa and A. Kawasaki, *J. Med. Chem.*, 1988, **31**, 84–91.
- 10 (a) E. Tsuji, K. Ando, J. Kunitomo, M. Yamashita, S. Ohta, S. Kohno and Y. Ohishi, Org. Biomol. Chem., 2003, 1, 3139–3141; (b) K. Ando, E. Tsuji, Y. Ando, N. Kuwata, J. Kunitomo, M. Yamashita, S. Ohta, S. Kohno and Y. Ohishi, Org. Biomol. Chem., 2004, 2, 625–635.
- 11 (a) P. D. Greenspan, A. J. Main, S. S. Bhagwat, L. I. Barsky, R. A. Doti, A. R. Eagle, L. M. Frey, H. Zhou, K. E. Lipson, M. H. Chin, R. H. Jackson and S. Uziel-Fusi, *Bioorg. Med. Chem. Lett.*, 1997, 7, 949–954; (b) P. D. Greenspan, R. A. Fujimoto, P. J. Marshall, A. Raychaudhuri, K. E. Lipson, H. Zhou, R. A. Doti, D. V. Coppa, L. Zhu, R. Pelletier, S. Uziel-Fusi, R. H. Jackson, M. H. Chin, B. L. Kotyuk and J. J. Fitt, *J. Med. Chem.*, 1999, **42**, 164–172.
- 12 M. Watanabe, S. Hisamatsu, H. Hotokezaka and S. Hurukawa, *Chem. Pharm. Bull.*, 1986, 34, 2810–2820.
- 13 (a) J. Boutagy and R. Thomas, *Chem. Rev*, 1974, 74, 87–99;
 (b) N. Matsuura, Y. Yashiki, S. Nakashima, M. Maeda and S. Sasaki, *Heterocycles*, 1999, 51, 975–978; (c) J. K. F. Geirsson, B. Ö. Gudmundsson and R. Sigurdardottir, *Acta Chem. Scand.*, 1993, 47, 1112–1116.
- 14 Y. Ohishi, T. Mukai, M. Nagahara, M. Yajima and N. Kajikawa, Chem. Pharm. Bull., 1989, 37, 2398–2404.
- 15 The torsion angle between the benzo[*b*]furan ring and the 2alkylcarbamoyl-1-methylvinyl group at the C-2 of (*E*)-**3a**, (*E*)-**3b** and (*E*)-**3f** was examined using Cambridge Soft, Chem3D 5.0, MM2. (*E*)-**3a**: 0.3°, (*E*)-3b: 3.5°, (*E*)-**3f**: 4.4°.
- 16 **3f**: Formula: C₁₇H₁₉NO₄, formula weight: 301.34, crystal system: monoclinic, space group: *P* 2₁/*a* (# 14), *a* = 7.309(1) Å, *b* = 15.546(1) Å, *c* = 13.8909(9), β = 104.340(1)°, *V* = 1529.1(3) Å³, *Z* = 4, *D*_{calc} = 1.309 g cm⁻³, *F*₀₀₀ = 640.00, μ(Cu-Kα) = 7.68 cm⁻¹, λ(Cu-Kα) = 1.54178 Å, ω-2θ scans at 23 °C, 2754 unique reflections (2θ < 135.1°), *R*1 = 0.066 [2240 reflections to calc. *R*1]. **4c**: Formula: C₃₁H₃₁NO₄, formula weight: 481.59, crystal system: triclinic, space group: *P*I (# 2), *a* = 102.59(1) Å, *b* = 16.120(2) Å, *c* = 8.654(2) Å, *a* = 104.45(1)°, β = 105.78(1)°, γ = 96.253(8)°, *V* = 1309.5(3) Å³, *Z* = 2, *D*_{calc} = 1.221 g cm⁻³, *F*₀₀₀ = 512.00, μ(Cu-Kα) = 6.42 cm⁻¹, λ(Cu-Kα) = 1.54178 Å, ω-2θ scans at

23 °C, 4727 unique reflections ($2\theta < 135.20^\circ$), R1 = 0.059 [2503 reflections to calc. R1]. CCDC reference numbers 245784 (**3f**) and 236333 (**4c**). See http://www.rsc.org/suppdata/ob/b4/b411286e/ for crystallographic data in .cif or other electronic format.

- 17 (a) F. R. Heck, Organic Reactions, John Wiley & Sons Publishers, New York, 1982, vol. 27, pp. 345–390; (b) P. D. Greenspan, R. A. Fujimoto, P. J. Marshall, A. Raychaudhuri, K. F. Lipson, H. Zhou, R. A. Doti, D. E. Coppa, L. Zhu, R. Pelletier, S. Uziel-Fusi, R. H. Jackson, M. H. Chin, B. L. Kotyuk and J. J. Fitt, J. Med. Chem., 1999, 42, 164–172.
- 18 (a) R. Adams and M. W. Miller, J. Am. Chem. Soc., 1940, 62, 53–57; (b) W. H. Mills and G. H. Dazeley, J. Chem. Soc., 1939, 460–463.
- 19 T. Nabe, H. Yamamura and S. Kohno, Jpn. J. Pharmacol., 1996, 70, 337–345.
- 20 Several methods, such as measurement of inhibition of human neutrophil LTB₄ receptor binding, LTB₄-induced neutrophil degradation, LTB₄-induced neutrophil chemotaxis, superoxide production by adherent human neutrophils, LTB₄- and fMLP-induced chemiluminescence, aggregation of human neutrophils, and cytoplasmic calcium concentrations upon LTB₄ stimulation of PMNLs, have been reported for in vitro assay of LTB4 antagonist in the following literature. (a) S. W. Djuric, P. W. Collins, P. H. Jones, R. L. Shone, B. S. Tosai, D. J. Fretland, G. M. Butchko, D. Villani-Prince, R. H. Keith, J. M. Zemaitis, L. Metcalf and R. F. Bauer, J. Med. Chem, 1989, 32, 1147-1156; (b) K. Koch, L. S. Melvin, Jr., L. A. Reiter, M. S. Biggers, H. J. Showell, R. J. Griffiths, E. R. Pettipher, J. B. Cheng, A. J. Milici, R. Breslow, M. J. Conklyn, M. A. Smith, B. C. Hackman, N. S. Doherty, E. Salter, C. A. Farrell and G. Schulte, J. Med. Chem, 1989, 37, 3197-3199; (c) J.-M. Poudrel, P. Hullot, J.-P. Girard, J.-C. Rossi, A. Muller, C. Bonne, V. Bezuglov, I. Serkov, P. Renard and B. Pfeiffer, J. Med. Chem., 1999, 42, 5289-5310.

²¹ These data are also reported as simple IC_{50} values.

Compound	IC_{50} for $hBLT_1/M$	IC ₅₀ for hBLT ₂ /M
ZK-158252 ONO-4057 4b (<i>E</i>)- 3a (<i>E</i>)- 3b	$5.4 \times 10^{-8} \\ 6.2 \times 10^{-6} \\ 4.2 \times 10^{-7} \\ > 10^{-5} \\ > 10^{-5}$	$\begin{array}{c} 3.1 \times 10^{-8} \\ 4.7 \times 10^{-6} \\ 4.8 \times 10^{-7} \\ > 10^{-5} \\ 8.3 \times 10^{-7} \end{array}$

- 22 Inhibition of calcium mobilization in CHO-hBLT₂ by (*E*)-**3b** was not due to its cytotoxicity, because this compound at 10 μ M did not affect calcium mobilization in CHO-hBLT₁ (Fig. 6). We also have confirmed that the inhibition by these compounds was not due to their cytotoxicity because they did not affect ATP-dependent calcium mobilization through intrinsic ATP receptors in CHO cells at the concentrations of BLT inhibition (data not shown).
- 23 H.-P. Nothacker, Z. Wang, Y. Zhu, R. K. Reinscheid, S. H. S. Lin and O. Civelli, *Mol. Pharmacol.*, 2000, 58, 1601–1608.
- 24 T. Yokomizo, T. Izumi, K. Chang, Y. Takuwa and T. Shimizu, *Nature*, 1997, **387**, 620–624.